

# Associations Between Gut Microbes and Social Behavior in Healthy 2-Year-Old Children

Desiree R. Delgadillo, MA, Sarah D. Pressman, PhD, Lisa M. Christian, PhD, Jeffrey D. Galley, PhD, and Michael T. Bailey, PhD

## ABSTRACT

**Objective:** Emerging research has connected abundances of specific bacteria to differences in psychosocial behaviors in animals and adult humans. However, research assessing mind-microbiome associations in children is sparse with extant work primarily focused on populations with autism, making it unclear whether links are also present in typically developing children. The current study fills this gap by examining associations between prosocial–self-regulating temperaments (effortful control; EC) and the gut microbiome in typically developing children.

**Methods:** Maternal ratings of temperament were assessed in 77 toddlers 18 to 27 months of age (46.7% female, mean age = 23.14 months). Next-generation pyrosequencing of the V1–V3 region of the 16S rRNA gene was used to classify children's gut microbial composition from fecal samples. EC included the following subcategories: cuddliness, attentional focusing, attentional shifting, inhibitory control, and low-intensity pleasure.

**Results:** After adjusting for covariates, EC was positively associated with relative abundances of Akkermansia ( $\Delta R^2 = 0.117$ ,  $b = 0.022$ ,  $SE = 0.007$ ,  $p = .002$ ), with cuddliness (i.e., joy and ease of being held) driving the relation. Furthermore, attentional focusing was negatively associated with *Alistipes* ( $\Delta R^2 = 0.062$ ,  $b = -0.011$ ,  $SE = 0.005$ ,  $p = .028$ ). Permutational analysis of variance revealed no significant differences in community structure between high and low EC groups on the phylum level ( $R^2 = 0.00372$ ,  $p = .745$ ) or the genus level ( $R^2 = 0.01559$ ,  $p = .276$ ).

**Conclusions:** Findings suggest that certain microbes may be linked to prosocial behaviors used to regulate emotion in typically developing children. Further research is needed to test whether these observations replicate in larger samples.

**Key words:** human microbiome, effortful control, prosocial behavior, microbial composition, preschool children.

## INTRODUCTION

Trillions of diverse microorganisms inhabit the human gut to form personalized microbial ecosystems. The gut microbiome is composed of bacteria, archaea, and eukaryotes and includes both pathogenic and symbiotic microbes that influence health outcomes (1). Interestingly, emerging evidence has revealed that bidirectional communication is taking place between the mind and the gut microbiome (2) as shown in numerous animal studies (3). Similarly, research on adults has echoed these connections and linked gut microbial composition to psychological traits and processes like the stress response and mood disorders (3,4), and to neurological conditions such as Parkinson disease (5). Importantly, recent work has also indicated that this may start at a young age, with important implications for both emotional and social functioning. However, childhood mind-microbiome literature is sparse and has primarily focused on patients with autism spectrum disorder (ASD), a condition characterized by atypical emotional, attentional, and social behaviors (6,7). Thus, little is known as to whether the microbiome is linked to adaptive psychological constructs in typically developing children. The current study will

contribute to this gap in the literature and explore whether certain microbes are connected to prosocial behaviors and constructs in healthy toddlers.

Given the robust body of research examining bidirectional mind-microbiome relations in animals, we first explore this more established research to inform the current study. Early work indicates that social factors can shape microbial composition. For instance, symbiotic bacteria (i.e., *Lactobacillus*) in infant rhesus monkeys significantly decreased after maternal separation (8). Furthermore, rodents placed with an aggressive cage mate resulted in decreased levels of the symbiotic bacteria *Bacteroides* and increased levels of the pathogenic bacteria *Clostridium* (9). However, numerous rodent studies have also shown that the gut microbiome regulates social behaviors (3). For instance, both rats and mice reared in the absence of microbial colonization (germ-free) showed decreased social approach behaviors when introduced to novel rodents compared with conventionally colonized controls (10,11). Similarly, germ-free mice chose to spend more

ASD = autism spectrum disorder, EC = effortful control, bTEFAP = bacterial tag-encoded FLX-amplicon pyrosequencing

## SDC Supplemental Digital Content

From the Department of Psychological Science (Delgadillo, Pressman), University of California Irvine, Irvine, California; Department of Psychiatry (Christian, Bailey), The Ohio State University, Columbus, Ohio; Institute for Behavioral Medicine Research, The Ohio State University, College of Medicine (Galley); and Abigail Wexner Research Institute at Nationwide Children's Hospital (Bailey), Columbus, Ohio.

Address correspondence to Desiree R. Delgadillo, MA, Department of Psychological Science, 4201 Social and Behavioral Sciences Gateway, University of California Irvine, Irvine, CA 92697-7085. E-mail: desic@uci.edu

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time in a solitary chamber over a social chamber and also showed abnormally low interest in unfamiliar mice compared with controls. Interestingly, microbial reconstitution helped to reverse some of the social deficits observed (12), providing preliminary evidence that certain social behaviors are linked to the gut microbiome. These studies point to the interesting possibility that microbial transmission is fostered by host sociality. Thus, some microbes may have evolved to promote positive social behaviors, for example, by decreasing anxiety (13) and encouraging prosocial behaviors such as the likelihood of exploration, interactions with novel individuals, and general increases in socialization (12).

Adult human studies complement the animal research by connecting microbial composition to psychological domains directly related to social behavior including stress responses, anxiety, and depression. For instance, a meta-analysis of seven human studies showed improvement in depression, anxiety, and perceived stress in participants who ingested a probiotic compared with those given a placebo (14). Similarly, depression is positively linked to naturally occurring levels of *Bacteroidales*, *Lachnospiraceae*, and *Alistipes* (15). Research has also found greater microbial diversity in those with larger social networks, and lower diversity in those with high anxiety and stress (typically tied to reduced sociability) (16). Intriguingly, the same pattern is found in married individuals, especially those with close relationships, who harbor microbial communities of greater diversity relative to those living alone (17).

Although sparse, research assessing mind-microbiome relations in healthy children is promising. For example, one study revealed links between happiness and microbial composition in a sample of typically developing children (18). Furthermore, a study of fifty-one 12-month-olds in China revealed associations between temperament and gut microbiota. Specifically, *Hungatella* (a member of the Clostridiaceae family) was negatively correlated with cuddliness, whereas the ability to be easily soothed was positively correlated with *Bifidobacterium* (19). Another study of healthy toddlers revealed a connection between extraverted temperaments and greater microbial diversity. This study also showed that, in girls only, higher EC is tied to bacterial diversity within samples (20). Together, this growing body of research suggests that certain features of socially adaptive psychological constructs, including temperament, are bidirectionally associated with microbial composition. Furthermore, there is a related body of work indicating that numerous environmental factors overlap to shape the microbiome from birth. From infancy to childhood, the composition of the gut microbiome becomes more complex and diverse, and these changes coincide with dietary changes such as weaning and the introduction of solid foods (21). Other factors known to shape the gut microbiome in toddlers are mode of delivery (vaginal versus cesarean delivery) (22), sex (23), body composition (24), and breastfeeding duration (15).

Although there is little work on social-microbiome connections in typically-developing children, we can draw from the more established child ASD literature to identify autism linked microbes and assess whether these bacteria are also related to emotion regulation and social behaviors among a group of typically developing children. Given evidence that social deficits characteristic of ASD are continuously distributed across the population (25,26), gut microbiome composition may also be related to variation in sociability in the general population. This body of work points to a

number of bacterial candidates. For example, young ASD patients have lower relative abundances of the bacterium *Dialister* (27), *Akkermansia* (28–30), and *Parabacteroides* (31) and higher relative abundances of *Lachnospiraceae* (32) and *Ruminococcus* (31,33) relative to healthy controls. Other bacteria frequently associated with ASD include *Bacteroides*, *Prevotella*, *Escherichia/Shigella*, and *Alistipes* (25,26). Specifically, research reveals increased levels of certain strains of *Bacteroides* in those with autism, whereas other research indicates no difference compared with healthy controls (25,27). Similarly, some studies found elevated levels of *Prevotella* (29,30), *Escherichia/Shigella* (34), and *Alistipes* (29), whereas other studies revealed decreased levels of *Prevotella* (35), *Escherichia/Shigella* (30), and *Alistipes* (34) when compared with controls. Taken together, these mixed findings suggest that certain bacteria are symbiotic with the host within optimal parameters, that is, not excessively high or low in relative abundance compared with typically developing individuals.

Indeed, early childhood is a crucial time in which the microbiome becomes established, and this coincides with a child's emerging temperament, making it a pivotal period in which to examine mind-microbiome connections. In the first years of life, researchers assess adaptive social temperaments via a child's ability to modify attention, emotion, and behavior (36). One example is the effortful control (EC) component of temperament, which is composed of subcategories assessing factors like child cuddliness (i.e., joy and ease of being held) and attentional focusing (i.e., ability to hold focus and resist distraction) (37). The cuddliness subcategory of EC is especially interesting in this context because it is the only EC category that effectively captures the features of many atypical socioemotional disorders (i.e., the ability/inability to engage in prolonged physical touch to emotion-regulate, presence/absence of prosocial behavior).

The current work uses data from a past study (20) assessing the relations between temperament and microbial composition in healthy toddlers. We build upon this research to examine whether microbiome characteristics typically tied to ASD-related behavior are associated with emotion regulation and social behavior in a sample of typically developing children. Bacteria assessed include *Bacteroides*, *Prevotella*, *Ruminococcaeaceae*, *Akkermansia* (38), *Escherichia/Shigella*, *Alistipes*, *Parabacteroides*, *Lachnospiraceae*, and *Dialister* (39) with a particular interest in the EC prosocial subcategory: cuddliness. We predict that higher levels of *Akkermansia*, *Dialister*, and *Parabacteroides* and lower levels of *Lachnospiraceae* and *Ruminococcus* will be linked to higher levels of EC. In light of the mixed literature regarding *Bacteroides*, *Prevotella*, *Escherichia/Shigella*, and *Alistipes*, we test these associations in an exploratory manner.

## METHODS

### Participants

Participants included 79 mother-child dyads recruited from the general community of Columbus, Ohio. This sample size was based on a power analysis with power set at 0.95 and effect size set at 0.2. Participants were excluded if the parent reported that the child had a developmental delay or a major health condition, or if the child was already toilet trained (because of sampling requirements). A more detailed report of recruitment of this sample is described by Christian and colleagues (20). Two microbial samples were removed because of low sequence count (<5108), leaving a final

sample of 77 toddlers for analyses (41 boys and 36 girls; mean [standard deviation], or M [SD] age = 23.14 [2.00] months). Mothers were 87.0% White ( $n = 67$ ), 9.1% Black ( $n = 7$ ), and 3.9% Asian ( $n = 3$ ). Mother's self-reported annual household income in US dollars was coded on a scale of 1 to 6, with 1 representing less than \$15,000 per year and 6 representing \$100,000 or higher. The average annual household income was \$50,000 to \$74,999, coded as 4 (M [SD] = 4.04 [1.62]). Parents' education level was coded on a scale from 1 to 7 (less than a seventh-grade education, junior high, some high school, high school graduate, some college, college graduate, some graduate school or higher). Mother's average education level was "college graduate," coded as a 6 (M [SD] = 6.09 [0.98]). Father's average education level was "some college," coded as a 5 (M [SD] = 5.65 [1.17]). Mean (SD) maternal age at the time of delivery was 31.1 (5.43) years, and 87.0% of mothers ( $n = 67$ ) were married. This study was approved by the Ohio State University Biomedical Institutional Review Board. Mothers provided informed written consent on behalf of themselves and their children and were compensated for participation. Data were collected between May 2011 and December 2012.

### Microbial Assessment

16S rRNA gene sequencing data, previously published by Christian and colleagues (20), were probed in the current study. Phylum and genus relative abundances are provided in Tables S1 and S2 (Supplemental Digital Content, <http://links.lww.com/PSYMED/A851>), and from these, a total of nine bacterial genera were selected for analysis based on previous literature commonly linking each genus with ASD. ASD-linked genera include the following: *Bacteroides*, *Prevotella*, *Ruminococcaeae*, *Akkermansia* (38), *Escherichia/Shigella*, *Alistipes*, *Parabacteroides*, *Lachnospiraceae*, and *Dialister* (39). The Benjamini-Hochberg method (40) was used to control for the false discovery rate. False discovery rates ( $q$  values) in genome and metagenome (microbiome) research vary and include  $q$  values such as 0.15 (41). In the current study, we considered a  $p$  value  $<.05$  and a  $q$  value  $<.15$  significant. Preliminary analyses revealed that *Akkermansia*, *Dialister*, and *Alistipes* were the only select bacterial genera significantly associated with EC and EC subcategories; thus, these three genera are the primary focus of the current study.

### Procedure

After providing consent, mothers completed online questionnaires describing the child's diet, body composition, duration of breastfeeding, demographic characteristics, and child temperament. Stool samples were collected from the toddler by the mother within 7 days of completing questionnaires.

### Collection and Storage of Stool Samples

Toddler stool samples were collected by mothers from children's diapers using sterile wooden applicators and 50-ml plastic conical collection tubes. Samples were refrigerated at approximately 4°C for up to 24 hours until delivery by research personnel or the participant to the Ohio State University Wexner Medical Center. Samples were transported in coolers with ice at which time samples were stored at -80°C until pyrosequencing was conducted.

### Measures

#### Effortful Control

EC was measured with a composite of five temperament subscales assessed by the short form of the Early Childhood Behavior Questionnaire (37). Mothers were asked to report the number of times in the last 2 weeks the child exhibited behavioral indicators of EC. Each subscale of EC consisted of six to eight items, and items were rated on a 7-point Likert scale (never, very rarely, less than half the time, half the time, more than half the time, almost always, always). Subscales included the following: attentional focusing, the toddler's ability to resist distraction and sustain focus on the object of concentration; attentional shifting; the ability to redirect focus from one target to another; cuddliness, the degree to which the child expresses

joy in and molds to the body of the caregiver when being held; inhibitory control, the ability to change behavior when instructed; and low-intensity pleasure, the level of pleasure or satisfaction derived from low stimulus activities that involve novelty, complexity, or incongruity. We examined each subscale for analyses and the composite score of all subscales to calculate overall EC. Specifically, associations between the 7-point Likert scale ratings for each subscale and the composite score of EC (composite scores ranged from 3.74 to 5.98) were assessed in relation to relative abundances of select bacteria.

#### Body Composition and Delivery Mode

Delivery mode (1, vaginal; 2, cesarean) and the child's sex (1, boys; 2, girls) were reported by mothers in the online questionnaire and included as covariates in analytic models. In addition, mothers reported the child's height and weight percentile from the most recent well-child visit to the pediatrician to assess body composition based on weight-to-height ratio. Weight-to-height ratios were converted to decimal numbers (Table 1) for statistical analyses.

#### Child Diet

Mothers completed an online questionnaire reporting the age in months in which fruits/vegetables, cereals/grains, and meats were introduced to the toddlers' diet and the frequency in which each food type was consumed. For analyses, food frequency item ratings were included as covariates. Specifically, mothers reported fruit/vegetable and meat consumption on a scale from 1 to 8 (less than once per month, once a month, once every 2 weeks, once a week, twice a week, every other day, once a day, two or more times per day). Mothers also reported on the occurrence and duration of breastfeeding (age in months in which breastfeeding stopped); duration was also included in analyses.

#### Bacterial Tag-Encoded FLX-Amplicon Pyrosequencing

Bacterial tag-encoded FLX-amplicon pyrosequencing (bTEFAP) was performed (42,43). The 16s rRNA universal primers 27f (AGA GTT TGA TCM TGG CTC AG) and 519r (GWATTACCGCGGCKGCTG) were used in a single-step 30-cycle polymerase chain reaction with the following thermoprofile: a single cycle of 94°C for 3 minutes, then 28 cycles of 30 seconds at 94°C, 40 seconds at 53°C, and 1 minute at 72°C, with a single 5-minute cycle at 72°C for 5 minutes for elongation. Amplicons were pooled at equivalent concentrations and purified (Agencourt Bioscience Corporation, Beverly, Massachusetts). Sequencing was performed with the Roche 454 FLX Titanium system using the manufacturer's guidelines.

#### Sequencing Analysis

The software package, Quantitative Insights Into Microbial Ecology (QIIME), version 1.8.0 (44), was used for filtering and analysis of attained sequences. Because the current study builds upon previous analyses that used QIIME, version 1.8.0, we continued with the use of this version so results were comparable. Quality filtering and demultiplexing were performed using the provided sequence file (.fasta) and sequence quality file (.qual). Filtering was completed with the following parameters: quality score  $>25$ , sequence length between 200 and 1000 bp, 6 allowed ambiguous bases, maximum of 6 homopolymer run, and zero allowed primer mismatches. On average, 14,862 sequences passed filtering per sample.

UClust (45) clustered sequences at 0.97 similarity into operational taxonomic units (OTUs). After representative sequence selection for each OTU, Greengenes v.13\_8 was used for taxonomic assignment (46). PyNAST was used for sequence alignment (44) with the Greengenes core reference alignment database (47). Sequences from boys and girls were filtered and de-multiplexed using the aforementioned method together, but were separated before OTU picking. Next-generation 454 pyrosequencing was used to identify bacterial communities. This approach was chosen owing to its low error rate and ability to classify microbes at lower taxonomic levels. Relative abundances of phylum and genera were used to assess relations between EC, community structure, and select bacterial genera.

**TABLE 1.** Descriptive Statistics of Key Variables by Children's Sex ( $n = 77$ )

Measures	Total ( $N = 77$ ), M (SD)	Boys ( $n = 41$ ), M (SD)	Girls ( $n = 36$ ), M (SD)	Sex Differences, $t$
Age, mo	23.14 (2.00)	23.20 (2.04)	23.08 (1.99)	-0.24
Body composition	0.93 (0.68)	1.04 (0.87)	0.80 (0.34)	-1.63
Breastfeeding duration	10.06 (7.23)	8.87 (6.34)	11.42 (8.00)	1.54
Vegetable/fruit consumption	7.00 (1.41)	6.76 (1.45)	7.28 (1.34)	1.63
Meat consumption	6.52 (1.80)	6.41 (1.73)	6.64 (1.85)	0.55
Effortful control	4.72 (0.47)	4.61 (0.40)	4.86 (0.52)	2.33
Cuddliness	4.36 (0.75)	4.27 (0.65)	4.46 (0.84)	1.10
Attentional focus	4.89 (0.75)	4.84 (0.68)	4.94 (0.83)	0.60
Attentional shifting	5.09 (0.69)	5.02 (0.69)	5.17 (0.68)	1.00
Inhibitory control	4.19 (0.96)	3.94 (0.91)	4.47 (0.95)	2.49*
Low-intensity pleasure	5.11 (0.77)	4.98 (0.71)	5.26 (0.81)	1.57
<i>Dialister</i>	0.06 (0.11)	0.07 (0.13)	0.06 (0.10)	-0.25
<i>Alistipes</i>	0.01 (0.03)	0.02 (0.04)	0.01 (0.02)	-0.76
<i>Akkermansia</i>	0.01 (0.05)	0.004 (0.01)	0.02 (0.07)	1.26*

M (SD) = mean (standard deviation).

Sex coding: 1 for boys, 2 for girls; body composition: weight/height ratio; breast feeding duration: age in months when breastfeeding stopped; vegetable/fruit/meat consumption: reported on a 1- to 8-point Likert scale (1, never; 8, two or more times per day); cuddliness, attentional focus, attentional shifting, inhibitory control, low-intensity pleasure: rated on a 7-point Likert scale (1, never; 8, always); effortful control: composite scores of all effortful control subscales, with values ranging from 3.74 to 5.98; bacterial genera (*Dialister*, *Alistipes*, and *Akkermansia*): relative abundance/percentage of each bacterium. Mothers were 87.0% White ( $n = 67$ ), 9.1% Black ( $n = 7$ ), and 3.9% Asian ( $n = 3$ ), and the mean (standard deviation) maternal age at the time of delivery was 31.1 (5.43) years.

\* $p < .05$ .

## Analytical Approach

Alpha diversity was measured with the Shannon Diversity Index (48), which assesses bacterial abundance (richness) and equalness of these abundances (evenness) using QIIME. Beta diversity analyses were conducted in R statistical software using the *adonis* function in the *vegan* package to generate Bray-Curtis matrices (49) and perform permutational analysis of variance and analysis of similarities (ANOSIMs; uses the  $R$  statistic to compare means of groups that use rank variables) between high and low EC groups. Differences in bacterial relative abundances derived from QIIME were assessed using version 25 of SPSS. If participants were missing data for any of the variables used in our analyses, their data were excluded. Male and female participants were very similar, with the exception of *Akkermansia* levels (Table 1). We first completed preliminary correlational analyses to test whether abundances of select genera were associated with EC, as well as the subscales of EC. Pearson correlation coefficients are reported in Table 2. Independent-samples  $t$  tests revealed that there were no significant sex differences in levels of cuddliness ( $p = .27$ ) or attentional focusing ( $p = .56$ ). Descriptive statistics for key variables, overall, and by child sex are reported in Table 1.

Preliminary analyses revealed associations between select bacterial genera and overall EC. To probe these findings further, we conducted a series of hierarchical regressions on the genera that correlated with EC and EC subscales while adjusting for sex, diet (frequency of fruit/vegetable and meat consumption), body composition (weight to height ratio), and breastfeeding duration (age in months in which breastfeeding stopped) to assess the relations between each subscale. Covariates were chosen based on prior research suggesting that sex (23), delivery mode (22), diet (50), body composition (24), and breastfeeding duration (51) influence gut microbial profiles. Although breastfeeding is linked to compositional changes in the microbiome (51), the mean age of the current sample is 23.14 months and microbial profiles resemble those found in adults by approximately 2 (52,53) to 3 years of age (54). Indeed, progression from infancy to childhood includes dietary changes such as weaning and an increase in solid food consumption, both of which impact microbial composition (21). Because our sample falls within this nutritional and developmental transition,

we conduct each analysis twice, once adjusting for breastfeeding duration and once excluding it from the statistical model. We report the adjusted  $R^2$  and  $p$  values in step 1 of each model, and the  $R^2$  change, unstandardized  $\beta$  coefficient, standard error, and  $p$  values in step 2 of each model.

## RESULTS

To determine whether EC was related to overall measures of microbial community composition, we conducted regression analyses between EC and diversity measures. Regression analyses revealed no significant association between levels of EC and alpha diversity ( $r = 0.123$ ,  $p = .293$ ). In addition, permutational analysis of variance analyses of Bray-Curtis (49) dissimilarities (used to quantify variation in genus and phylum between samples) revealed that there were no significant differences in community structure between high and low EC groups on the phylum level ( $R^2 = 0.00372$ ,  $p = .745$ ; Figure 1) or on the genus level ( $R^2 = 0.01559$ ,  $p = .276$ ; see Figure 2). ANOSIM analyses of Bray-Curtis (49) dissimilarities were also conducted. ANOSIM's  $R$  statistic measures dissimilarities between groups for rank variables (i.e., relative abundances) by comparing within a between-group differences (55). Results of the ANOSIM revealed no significant differences between high and low EC groups on the phylum ( $R = -0.0002$ ,  $p = .545$ ) or on the genus level ( $R = -0.012$ ,  $p = .818$ ). Relative abundance data for phylum and genera are reported in Supplemental Tables S1 and S2, <http://links.lww.com/PSYMED/A851>.

Despite the lack of relationships between EC and measures of microbial diversity, EC was related to the relative abundances of three predicted bacterial taxa. Based on our a priori hypotheses, zero-order correlations among EC and select bacterial taxa (presented in Table 2) revealed that EC was significantly positively intercorrelated with *Akkermansia* ( $r(75) = 0.25$ ,  $p = .027$ ) and *Dialister* ( $r(75) = 0.24$ ,  $p = .034$ ) and negatively correlated with

**TABLE 2.** Correlation Matrix for Key Variables ( $n = 77$ )

Variables	1	2	3	4	5	6	7	8	9
1. Body composition	—								
2. Breastfeeding duration	-0.01	—							
3. Vegetable/fruit consumption	-0.20	-0.15	—						
4. Meat consumption	0.06	-0.11	0.41**	—					
5. Effortful control	-0.08	-0.10	0.07	0.12	—				
6. Cuddliness	0.04	-0.08	0.01	0.08	0.65**	—			
7. Attentional focus	0.06	-0.02	0.10	0.07	0.52**	0.07	—		
8. <i>Dialister</i>	-0.08	0.04	-0.03	0.00	0.24*	0.23*	0.16	—	
9. <i>Alistipes</i>	0.14	-0.03	-0.11	-0.25*	-0.24*	-0.02	-0.27*	0.02	—
10. <i>Akkermansia</i>	0.02	0.11	-0.14	-0.03	0.25*	0.36**	0.00	0.31**	-0.07

Sex coding: 1 for boys, 2 for girls.

\* $p < .05$ .

\*\* $p < .01$ .

*Alistipes* ( $r(75) = -0.24, p = .035$ ). Follow-up analyses of EC's five subscales revealed that significant findings were driven by cuddliness and attentional focusing. Specifically, cuddliness was positively associated with *Akkermansia* ( $r(75) = 0.36, p = .001$ ) and *Dialister* ( $r(75) = 0.23, p = .049$ ). In addition, attentional focusing was negatively associated with *Alistipes* ( $r(75) = -0.27, p = .017$ ).

#### Association Between Cuddliness and *Akkermansia*

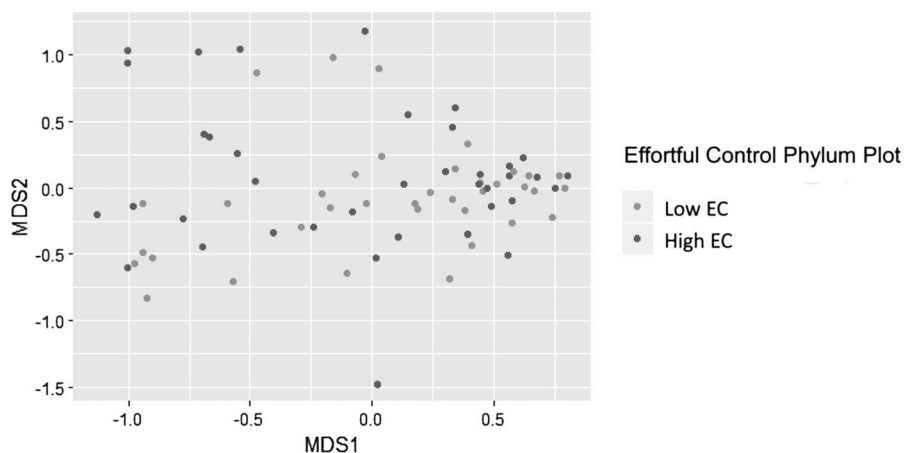
After adjusting for sex, delivery route, diet, body composition, and breastfeeding duration (adjusted  $R^2 = -0.024, p = .646$ ), cuddliness was positively associated with higher levels of *Akkermansia* ( $\Delta R^2 = 0.116, b = 0.022, SE = 0.007, p = .003$ ). Next, we adjusted for sex, delivery route, diet, and body composition but excluded breastfeeding duration from the model (adjusted  $R^2 = -0.014, p = .558$ ). Cuddliness remained positively associated with higher levels of *Akkermansia* ( $\Delta R^2 = 0.117, b = 0.022, SE = 0.007, p = .002$ ).

#### Association Between Cuddliness and *Dialister*

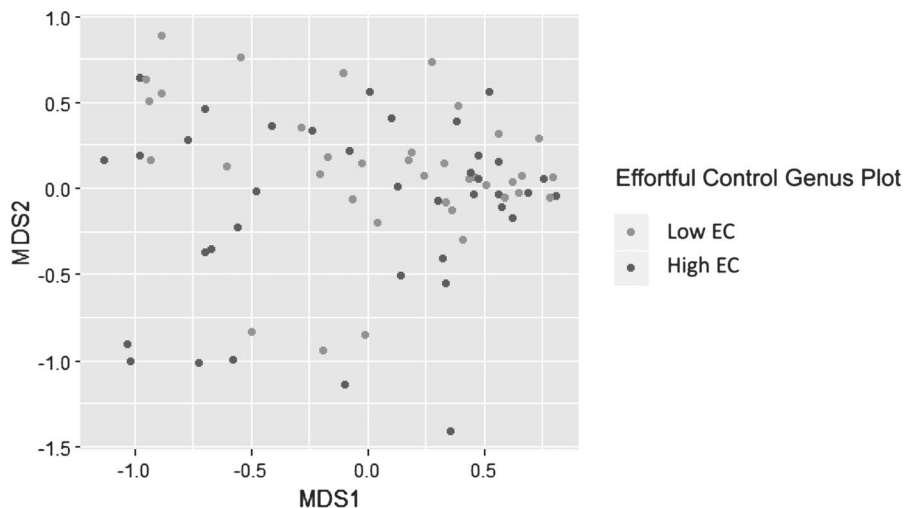
We then probed the relation between cuddliness and *Dialister* while adjusting for sex, delivery route, diet, body composition, and breastfeeding duration (adjusted  $R^2 = -0.75, p = .994$ ). Cuddliness was no longer associated with higher levels of *Dialister* in step 2 of the model ( $\Delta R^2 = 0.053, b = 0.036, SE = 0.018, p = .052$ ). Next, we conducted the analysis again using the same covariates but excluded breastfeeding duration (adjusted  $R^2 = -0.065, p = .997$ ). Cuddliness was still not significantly associated with *Dialister* in step 2 of the model ( $\Delta R^2 = 0.052, b = 0.035, SE = 0.018, p = .054$ ) at an  $\alpha$  level of .05.

#### Association Between Attentional Focusing and *Alistipes*

Finally, we conducted another series of hierarchical regressions to examine the association between attentional focusing and the relative abundance of *Alistipes*. Results of the analyses revealed that after accounting for covariates (covariate adjusted  $R^2 = -0.004, p = .463$ ), attentional focusing was negatively associated with



**FIGURE 1.** NMDS ordination using Bray-Curtis dissimilarity to calculate distances between samples on the phylum level. EC = effortful control; NMDS = nonmetric multidimensional scaling.



**FIGURE 2.** NMDS ordination using Bray-Curtis dissimilarity to calculate distances between samples on the genus level. EC = effortful control; NMDS = nonmetric multidimensional scaling.

levels of *Alistipes* ( $\Delta R^2 = 0.061$ ,  $b = -0.011$ ,  $SE = 0.005$ ,  $p = .031$ ). These results remained significant after, excluding breastfeeding duration from the model (covariate adjusted  $R^2 = 0.005$ ,  $p = .377$ ). Attentional focusing was again negatively associated with levels of *Alistipes* ( $\Delta R^2 = 0.062$ ,  $b = -0.011$ ,  $SE = 0.005$ ,  $p = .028$ ).

## DISCUSSION

The current study showed that *Akkermansia* and *Dialister* were positively associated with EC and Cuddliness. In addition, *Alistipes* was negatively related to attentional focusing. These findings complement past mind-microbiome research (27,28) and indicate that the microbiome is connected to social behaviors in a healthy, young, toddler sample. One possible explanation for links between *Akkermansia*, EC, and cuddliness may be the bacterium's role in stimulating the production of mucous that lines the gut (56). Evidence suggests that low levels of *Akkermansia* are associated with compromised mucous barrier function resulting in the displacement of gut microbiota and/or their metabolic products into the serum (i.e., leaky gut) and heightened inflammation (28). This is thought to be a mechanism connecting the microbiome to mood and neurological disorders (57). In addition, the current study also showed positive associations between *Dialister* and cuddliness, which supports our hypotheses and is in line with past work linking lower levels of *Dialister* to depression, anxiety (58), and ASD (34), three conditions commonly marked by social deficits. Interestingly, cuddliness is the only subcategory of EC that exclusively uses prosocial behavior as a means by which children regulate emotion. Thus, one possibility for these mind-microbiome associations is that *Akkermansia* and *Dialister* promote social behaviors in humans; however, future experimental and longitudinal research is needed to test this suggestion. Finally, given the connection between *Alistipes* in tryptophan metabolism (a precursor of serotonin) (59), there is a mechanistic reason to support this bacterial group playing a role in mood-related functioning, which is also supported by past work connecting *Alistipes* levels to ASD symptoms (29).

Contrary to our hypothesis, the remaining six bacterial taxa were not related to adaptive host behaviors in this sample. It is pos-

sible that, of the select nine bacteria that we chose to test, the non-significant six only influence psychological processes on the disease-disorder end of the continuum. In addition, much of the research on ASD-microbiome links is conducted among a wide age range of youth aged 2 to 18 years. It could be that the inverse associations we predicted would not be evident until later into childhood or adolescence.

This study is among the first to examine whether there is a prosocial, self-regulating counterpart to ASD behavior-microbiome associations in typically developing toddlers. Existing research examining mind-microbiome relations is primarily geared to investigate pathology. Although understanding pathological associations is vitally important, the subsequent interventions or therapies developed would only be designed to treat those exhibiting symptoms at a diagnosable severity. Clearly, the majority of the population is not diagnosed with ASD; yet, many typically developing individuals also experience some level of suffering linked to problems with self-regulation and prosocial behaviors. The current study may guide the development of evidence-based interventions that improves EC for both typical and atypical children alike. That is, future researchers could build upon this work to discover whether it is possible to calibrate the gut microbiome to promote emotion regulation and sociality, two features of well-being that affect the quality of life across the life span, regardless of any diagnoses. If studies in larger cohorts replicate these findings, possible interventions could include the use of a probiotic containing *Akkermansia* or dietary interventions that promote the growth of *Akkermansia*. This work may also help to set parameters of what constitutes an "optimal range" of *Akkermansia* and *Alistipes* because research has still not identified what defines excessively high versus low relative abundances of this bacterium in relation to well-being.

## Limitations

The current study has many strengths; however, it is important to note that this work is limited by cross-sectional design, self-reporting of predictor variables, and a lack of ethnic diversity. Furthermore, the frequency of physical contact with parents, siblings,

other children, and microbial composition of the home environment was not measured but may have influenced results. In addition, the current study used multiple comparison procedures to minimize type I error; however, replication work in larger cohorts is needed to undergird these findings. Finally, although research on the relations between a single bacterial taxa and psychological processes has revealed important mind-gut connections, future researchers should also consider how these associations may vary in the presence and abundance of other microbes. Thus, we underscore the need for future research examining relations between EC and microbial composition with the aforementioned factors considered.

Emerging findings linking gut microbes to social processes in humans open numerous avenues for further inquiry. Future work should further consider possible origins of bacterial population differences (e.g., hygiene hypothesis, overexposure to antibiotics at an early age, microbial composition of the home/family environment) and how this translates into later differences in social processes. The gut microbiome has been well described as an ecosystem, and as such, the stability of this ecosystem may be influenced by the presence or abundance of cohabitating organisms, an individual's gene expression, and the environment in which the host lives. Each of these variables could influence the stability of the gut microbial community. As this field advances, the challenge for future researchers will be to design methodologies that capture these dynamic and sensitive systems. Despite challenges, emerging research linking microbes to social behavior is promising and opens a myriad of possible ways in which typical and atypical psychological processes may improve. Studies like this may illuminate numerous pathways by which microbes are linked to other highly adaptive psychological constructs in healthy children.

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## REFERENCES

- Shreiner AB, Kao JY, Young VB. The gut microbiome in health and in disease. *Curr Opin Gastroenterol* 2015;31:69–75.
- Mayer EA. Gut feelings: the emerging biology of gut-brain communication. *Nat Rev Neurosci* 2011;12:453–66.
- Archie EA, Tung J. Social behavior and the microbiome. *Curr Opin Behav Sci* 2015;6:28–34.
- Kelly JR, Kennedy PJ, Cryan JF, Dinan TG, Clarke G, Hyland NP. Breaking down the barriers: the gut microbiome, intestinal permeability and stress-related psychiatric disorders. *Front Cell Neurosci* 2015;9.
- Ghaisas S, Maher J, Kanthasamy A. Gut microbiome in health and disease: linking the microbiome-gut-brain axis and environmental factors in the pathogenesis of systemic and neurodegenerative diseases. *Pharmacol Ther* 2016;158:52–62.
- American Psychiatric Association. *Diagnostic and Statistical Manual of Mental Disorders*. Arlington, VA: American Psychiatric Association; 2013.
- Ashwood P, Wills S, Van de Water J. The immune response in autism: a new frontier for autism research. *J Leukoc Biol*. [Internet] 2006;80:1–15. doi:10.1189/jlb.1205707.
- Bailey MT, Coe CL. Maternal separation disrupts the integrity of the intestinal microflora in infant rhesus monkeys. *Dev Psychobiol* 1999;35:146–55.
- Bailey MT, Dowd SE, Galley JD, Huffnagle AR, Allen RG, Lyte M. Exposure to a social stressor alters the structure of the intestinal microbiota: implications for stressor-induced immunomodulation. *Brain Behav Immun* 2011;25:397–407.
- Arentsen T, Raith H, Qian Y, Forssberg H, Heijtz RD. Host microbiota modulates development of social preference in mice. *Microb Ecol Health Dis* 2015;26:29719.
- Crumeyrolle-Arias M, Jaglin M, Bruneau A, Vancassel S, Cardona A, Dauge V, Naudon L, Rabot S. Absence of the gut microbiota enhances anxiety-like behavior and neuroendocrine response to acute stress in rats. *Psychoneuroendocrinology* 2014;42:207–17.
- Desbonnet L, Clarke G, Shanahan F, Dinan TG, Cryan JF. Microbiota is essential for social development in the mouse. *Mol Psychiatry* 2014;19:146–8.
- Messaoudi M, Lalonde R, Violle N, Javelot H, Desor D, Nejdí A, Bisson J-F, Rougeot C, Pichelin M, Cazaubiel M, Cazaubiel J-M. Assessment of psychotropic-like properties of a probiotic formulation (*Lactobacillus helveticus* R0052 and *Bifidobacterium longum* R0175) in rats and human subjects. *Br J Nutr* 2011;105:755–64.
- McKean J, Naug H, Nikbakht E, Amiet B, Colson N. Probiotics and subclinical psychological symptoms in healthy participants: a systematic review and meta-analysis. *J Altern Complement Med* 2017;23:249–58.
- Naseribafrouei A, Hestad K, Avershina E, Sekelja M, Linlökken A, Wilson R, Rudi K. Correlation between the human fecal microbiota and depression. *Neurogastroenterol Motil* 2014;26:1155–62.
- Johnson KV-A. Gut microbiome composition and diversity are related to human personality traits. *Hum Microb J* 2020;15:100069.
- Dill-McFarland KA, Tang Z-Z, Kemis JH, Kerby RL, Chen G, Palloni A, Sorenson T, Rey FE, Herd P. Close social relationships correlate with human gut microbiota composition. *Sci Rep* 2019;9:703.
- Michels N, Van de Wiele T, Fouhy F, O'Mahony S, Clarke G, Keane J. Gut microbiome patterns depending on children's psychosocial stress: reports versus biomarkers. *Brain Behav Immun* 2019;80:751–62.
- Wang Y, Chen X, Yu Y, Liu Y, Zhang Q, Bai J. Association between gut microbiota and infant's temperament in the first year of life in a Chinese birth cohort. *Microorganisms* 2020;8:753.
- Christian LM, Galley JD, Hade EM, Schoppe-Sullivan S, Kamp Dush C, Bailey MT. Gut microbiome composition is associated with temperament during early childhood. *Brain Behav Immun* 2015;45:118–27.
- Ku H-J, Kim Y-T, Lee J-H. Microbiome study of initial gut microbiota from newborn infants to children reveals that diet determines its compositional development. *J Microbiol Biotechnol* 2020;30:1067–71.
- Reyman M, van Houten MA, van Baarle D, Bosch AATM, Man WH, Chu MLJN, Arp K, Watson RL, Sanders EAM, Fuentes S, Bogaert D. Impact of delivery mode-associated gut microbiota dynamics on health in the first year of life. *Nat Commun* 2019;10:4997.
- Jašarević E, Morrison KE, Bale TL. Sex differences in the gut microbiome-brain axis across the lifespan. *Philos Trans R Soc Lond B Biol Sci* 2016;371:20150122.
- O'Sullivan A, Farver M, Smilowitz JT. The influence of early infant-feeding practices on the intestinal microbiome and body composition in infants. *Nutr Metab Insights* 2015;8(Suppl 1):1–9.
- Constantino JN, Todd RD. Autistic traits in the general population. *Arch Gen Psychiatry* 2003;60:524.
- Ruzich E, Allison C, Smith P, Watson P, Auyeung B, Ring H, Baron-Cohen S. Measuring autistic traits in the general population: a systematic review of the Autism-Spectrum Quotient (AQ) in a nonclinical population sample of 6,900 typical adult males and females. *Mol Autism* 2015;6:2.
- Liu F, Li J, Wu F, Zheng H, Peng Q, Zhou H. Altered composition and function of intestinal microbiota in autism spectrum disorders: a systematic review. *Transl Psychiatry* 2019;9:43.
- Wang L, Christophersen CT, Sorich MJ, Gerber JP, Angley MT, Conlon MA. Low relative abundances of the mucolytic bacterium *Akkermansia muciniphila* and *Bifidobacterium* spp. in feces of children with autism. *Appl Environ Microbiol* 2011;77:6718–21.
- De Angelis M, Piccolo M, Vannini L, Siragusa S, De Giacomo A, Serrazanetti DI, Cristofori F, Guerzoni ME, Gobetti M, Francavilla R. Fecal microbiota and metabolome of children with autism and pervasive developmental disorder not otherwise specified. *PLoS One* 2013;8:e76993.
- Zou R, Xu F, Wang Y, Duan M, Guo M, Zhang Q, Zhao H, Zheng H. Changes in the gut microbiota of children with autism spectrum disorder. *Autism Res* 2020;13:1614–25.
- Xu M, Xu X, Li J, Li F. Association between gut microbiota and autism spectrum disorder: a systematic review and meta-analysis. *Front Psych* 2019;10.
- Ding X, Xu Y, Zhang X, Zhang L, Duan G, Song C, Li Z, Yang Y, Wang Y, Wang X, Zhu C. Gut microbiota changes in patients with autism spectrum disorders. *J Psychiatr Res* 2020;129:149–59.

33. Wang L, Christophersen CT, Sorich MJ, Gerber JP, Angley MT, Conlon MA. Increased abundance of *Sutterella* spp. and *Ruminococcus torques* in feces of children with autism spectrum disorder. *Mol Autism* 2013;4:42.
34. Strati F, Cavalieri D, Albanese D, De Felice C, Donati C, Hayek J, Jousson O, Leoncini S, Renzi D, Calabrò A, De Filippo C. New evidences on the altered gut microbiota in autism spectrum disorders. *Microbiome* 2017;5:24.
35. Andreo-Martínez P, García-Martínez N, Sánchez-Samper EP, Martínez-González AE. An approach to gut microbiota profile in children with autism spectrum disorder. *Environ Microbiol Rep* 2020;12:115–35.
36. Kochanska G, Murray KT, Harlan ET. Effortful control in early childhood: continuity and change, antecedents, and implications for social development. *Dev Psychol* 2000;36:220–32.
37. Putnam SP, Gartstein MA, Rothbart MK. Measurement of fine-grained aspects of toddler temperament: the Early Childhood Behavior Questionnaire. *Infant Behav Dev* 2006;29:386–401.
38. Ding HT, Taur Y, Walkup JT. Gut microbiota and autism: key concepts and findings. *J Autism Dev Disord* 2017;47:480–9.
39. Hughes HK, Rose D, Ashwood P. The gut microbiota and dysbiosis in autism spectrum disorders. *Curr Neurol Neurosci Rep* 2018;18:81.
40. Benjamini Y, Hochberg Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing. *J R Stat Soc B Methodol* 1995;57:289–300.
41. Cole CB, Nikpay M, Lau P, Stewart AFR, Davies RW, Wells GA, Dent R, McPherson R. Adiposity significantly modifies genetic risk for dyslipidemia. *J Lipid Res* 2014;55:2416–22.
42. Dowd SE, Callaway TR, Wolcott RD, Sun Y, McKeehan T, Hagevoort RG, Edrington TS. Evaluation of the bacterial diversity in the feces of cattle using 16S rDNA bacterial tag-encoded FLX amplicon pyrosequencing (bTEFAP). *BMC Microbiol* 2008;8:125.
43. Dowd SE, Wolcott RD, Sun Y, McKeehan T, Smith E, Rhoads D. Polymicrobial nature of chronic diabetic foot ulcer biofilm infections determined using bacterial tag encoded FLX amplicon pyrosequencing (bTEFAP). *PLoS One* 2008;3:e3326.
44. Caporaso JG, Kuczynski J, Stombaugh J, Bittinger K, Bushman FD, Costello EK, Fierer N, Peña AG, Goodrich JK, Gordon JL, Huttley GA, Kelley ST, Knights D, Koenig JE, Ley RE, Lozupone CA, McDonald D, Muegge BD, Pirrung M, Reeder J, Sevinsky JR, Turnbaugh PJ, Walters WA, Widmann J, Yatsunenko T, Zaneveld J, Knight R. QIIME allows analysis of high-throughput community sequencing data. *Nat Methods* 2010;7:335–6.
45. Edgar RC. Search and clustering orders of magnitude faster than BLAST. *Bioinformatics* 2010;26:2460–1.
46. McDonald D, Price MN, Goodrich J, Nawrocki EP, DeSantis TZ, Probst A, Andersen GL, Knight R, Hugenholtz P. An improved Greengenes taxonomy with explicit ranks for ecological and evolutionary analyses of bacteria and archaea. *ISME J* 2012;6:610–8.
47. DeSantis TZ, Hugenholtz P, Larsen N, Rojas M, Brodie EL, Keller K, Huber T, Dalevi D, Hu P, Andersen GL. Greengenes, a chimera-checked 16S rRNA gene database and workbench compatible with ARB. *Appl Environ Microbiol* 2006;72:5069–72.
48. Shannon CE. The mathematical theory of communication. 1963. *MD Comput* 1997;14:306–17.
49. Bray JR, Curtis JT. An ordination of the upland forest communities of southern Wisconsin. *Ecol Monogr* 1957;27:325–49.
50. David LA, Maurice CF, Carmody RN, Gootenberg DB, Button JE, Wolfe BE, Ling AV, Devlin AS, Vanma Y, Fischbach MA, Biddinger SB, Dutton RJ, Turnbaugh PJ. Diet rapidly and reproducibly alters the human gut microbiome. *Nature* 2014;505:559–63.
51. Kashtanova DA, Popenko AS, Tkacheva ON, Tyakht AB, Alexeev DG, Boytsov SA. Association between the gut microbiota and diet: fetal life, early childhood, and further life. *Nutrition* 2016;32:620–7.
52. Koenig JE, Spor A, Scalfone N, Fricker AD, Stombaugh J, Knight R, Angenent LT, Ley RE. Succession of microbial consortia in the developing infant gut microbiome. *Proc Natl Acad Sci* 2011;108:4578–85.
53. Palmer C, Bik EM, DiGiulio DB, Relman DA, Brown PO. Development of the human infant intestinal microbiota. *PLoS Biol* 2007;5:e177.
54. Arrieta M-C, Stiemsma LT, Amenyogbe N, Brown EM, Finlay B. The intestinal microbiome in early life: health and disease. *Front Immunol* 2014;5:427.
55. Clarke KR, Somerfield PJ, Gorley RN. Testing of null hypotheses in exploratory community analyses: similarity profiles and biota-environment linkage. *J Exp Mar Biol Ecol* 2008;366:56–69.
56. Zhou K. Strategies to promote abundance of *Akkermansia muciniphila*, an emerging probiotics in the gut, evidence from dietary intervention studies. *J Funct Foods* 2017;33:194–201.
57. Maes M, Kubera M, Leunis J-C. The gut-brain barrier in major depression: intestinal mucosal dysfunction with an increased translocation of LPS from gram negative enterobacteria (leaky gut) plays a role in the inflammatory pathophysiology of depression. *Neuro Endocrinol Lett* 2008;29:117–24.
58. Taylor AM, Thompson SV, Edwards CG, MUSAAD SMA, Khan NA, Holscher HD. Associations among diet, the gastrointestinal microbiota, and negative emotional states in adults. *Nutr Neurosci* 2020;23:983–92.
59. de Theije CGM, Wopereis H, Ramadan M, van Eijndhoven T, Lambert J, Knol J, Garssen J, Kraneveld AD, Oozeer R. Altered gut microbiota and activity in a murine model of autism spectrum disorders. *Brain Behav Immun* 2014;37:197–206.